

dd-Check STEC Solution

A Multipronged Approach to STEC Testing

ddPCR Food Applications

- Microbial enumeration
- Virulence co-detection
- Food authenticity
- Parasites and viruses:
Non-cultural targets



Current quantitative PCR (qPCR)-based methods for screening Shiga toxin-producing *Escherichia coli* (STEC) make it challenging to differentiate between samples in which a single organism contains both *stx* and *eae* genes (true positive, linked virulence) and samples in which *stx* and *eae* reside in different organisms (false positive, unlinked virulence).

Droplet Digital™ PCR (ddPCR™) technology demonstrates the capacity of virulence linkage analysis by partitioning intact cells into droplets in which cell lysis and PCR amplification occur, enhancing the screening accuracy by reducing false-positive reactions. With a high level of agreement in STEC confirmation between the dd-Check STEC Solution and the U.S. Department of Agriculture (USDA) MLG 5C.01 method, the dd-Check STEC Solution proves useful for culture-independent confirmation following primary screening by qPCR assays.

A. Screening Method



B. Confirmatory Method



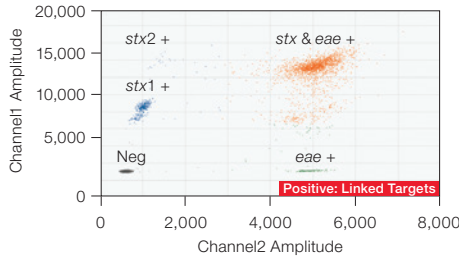
Fig. 1. Timelines for utilizing dd-Check STEC Solution as a screening and confirmatory method. Follow the timeline for using ddPCR as a primary screening assay (A) and as a confirmatory assay (B).

* Presumptive positive samples from iQ-Check STEC VirX PCR Detection Kit should be tested on iQ-Check STEC SerO II Kit if serotype identification is required.

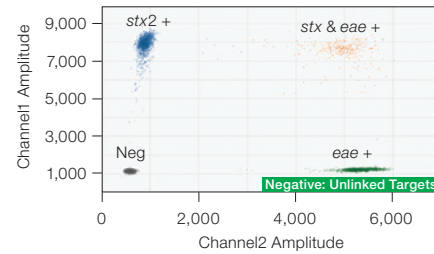
** Perform regrowth only if different media was used for screening or if iQ-Check STEC VirX Kit Cq is >34.



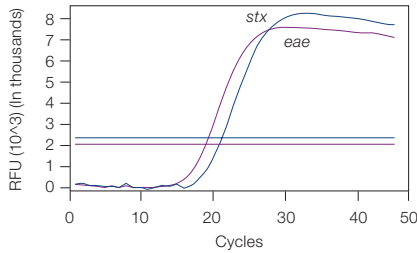
A. O157:H7 (linked *stx* and *eae*), tested with dd-Check STEC Solution



B. O45:H18 (*stx*+ *eae*-) and O45:H2 (*stx*- *eae*+), tested with dd-Check STEC Solution



C. O157:H7 (linked *stx* and *eae*), tested with iQ-Check STEC VirX Kit



D. O45:H18 (*stx*+ *eae*-) and O45:H2 (*stx*- *eae*+), tested with iQ-Check STEC VirX Kit

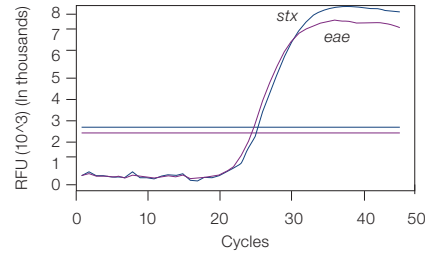


Fig. 2. Comparison of data output between ddPCR and qPCR in distinguishing samples with linked and unlinked virulence targets. This figure demonstrates that compared to the challenge of qPCR-based methods in differentiating sample enrichment with linked and unlinked targets, ddPCR technology was able to individually identify the presence of *stx* and *eae* to verify the co-existence of these two virulence genes via linkage analysis.

Research Studies Evaluating dd-Check STEC as a Culture-Independent Confirmation Method for STEC in Beef Samples

Fresh and frozen enrichments from presumptive positive samples were processed for culture-independent confirmation using the dd-Check STEC Solution and cultural detection and confirmation following the USDA MLG 5C.01.

Study 1: A major U.S. third-party food lab

Study Objects: 100 beef samples (beef trim and MicroTally) from a major beef producer in the U.S. that tested presumptive for *E. coli* O157:H7 via qPCR-based primary screening tests

Key Results

- For the confirmation of linked virulence profiles, the ddPCR assay and the USDA MLG 5C.01 identified the presence of linked targets in 97 and 95 samples, respectively, with 98% agreement
- Two of the four samples that were positive with the dd-Check STEC Solution but negative with MLG 5C.01 still produced colonies of STEC with *stx* and *eae*, but were negative with serogroup screening (Table 1)

Table 1. Samples that tested positive and negative for STEC and MLG.

	Positive for MLG	Negative for MLG
Positive for dd-Check STEC	93	4
Negative for dd-Check STEC	0	3

Study 2: A government research lab

Study Objects: 100 beef samples (beef trim and MicroTally) that tested presumptive for *stx* and *eae* via iQ-Check STEC VirX Kit and one of the top 7 serogroups via iQ-Check STEC SerO II Kit

Key Results

- For the confirmation of linked virulence profiles, the ddPCR assay identified the presence of a linked target in the four samples that were initially culturally confirmed by the USDA Food Safety and Inspection Service (FSIS)
- Four samples that were positive with the dd-Check STEC Solution but negative with MLG 5C.01 still produced colonies of STEC with linked *stx* and *eae*, but were negative with serogroup screening (Table 2)

Table 2. Samples that tested positive and negative for STEC and MLG.

	Positive for MLG	Negative for MLG
Positive for dd-Check STEC	4	4
Negative for dd-Check STEC	0	92

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