



# CERTIFICATION

## AOAC Research Institute *Performance Tested Methods*<sup>SM</sup>

Certificate No.  
**080701**

The AOAC Research Institute hereby certifies the method known as:

### **RAPID'*Listeria* spp. Agar**

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**Hercules, CA 94547**  
**USA**

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A handwritten signature in black ink that reads 'Scott Coates'.

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Scott Coates, Senior Director  
Signature for AOAC Research Institute

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<b>METHOD NAME</b> RAPID' <i>Listeria</i> spp. Agar		<b>CATALOG NUMBERS</b> 356-4744, 356-4745, 356-4746	
<b>INDEPENDENT LABORATORY</b> rtech Laboratories 4001 Lexington Ave. North Arden Hills, MN 55112			
<b>APPLICABILITY OF METHOD</b> Target organism – <i>Listeria</i> spp.  Matrixes – stainless steel, plastic, ceramic, and sealed concrete  Performance claims – RAPID' <i>Listeria</i> spp is a chromogenic medium for isolation and detection of <i>Listeria</i> spp from environmental surfaces.		<b>REFERENCE METHOD</b> <i>Microbiology Laboratory Guidebook</i> (October 1, 2004) U.S. Department of Agriculture, Food Safety and Inspection Service, Office of Public Health Science, Chapter 8.05. (2)	
<b>ORIGINAL CERTIFICATION DATE</b> August 15, 2007		<b>CERTIFICATION RENEWAL RECORD</b> Renewed annually through December 2024.	
<b>METHOD MODIFICATION RECORD</b> 1. January 2020 Level 1 2. January 2020 Level 1 3. October 2021 Level 1  4. October 2021 Level 1		<b>SUMMARY OF MODIFICATION</b> 1. Editorial/clerical changes and reformatting of insert. 2. Editorial/clerical changes. 3. Editorial changes and addition of user information in French, German, Spanish, Portuguese, and Italian. 4. Editorial/clerical changes.	
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#### PRINCIPLE OF THE METHOD (1)

RAPID' *Listeria* spp is a selective chromogenic plating medium for the isolation and the presumptive identification of all species of *Listeria*. The cultural properties of the medium are based on a balance of carefully selected growth-promoting nutrients, enzymatic enhancers and a mixture of selective agents, including lithium chloride and nalidixic acid. The presumptive chromogenic identification system relies on a chromogenic substrate that allows the detection of the  $\beta$ -D-glucosidase activity, an enzyme common to all species of *Listeria*. The hydrolysis of the substrate leads to the formation of a colored precipitate and all the presumptive *Listeria* positive colonies are blue. Combined with this principle, RAPID' *Listeria* spp utilizes an original selective mixture which enables the inhibition of most background flora. The few non-*Listeria* bacteria that do grow on the agar will produce colorless colonies.

#### DISCUSSION OF THE VALIDATION STUDY (1)

Chromogenic media rely on the enzymatic activity of certain target organisms. The chromogens are selected based on the properties of these specific target organisms. These specific compounds, in addition to a selective mixture, make these media selective and specific. The use of chromogenic media has been demonstrated in previous studies (3,4). This highly selective media can reduce time to results. In this validation study, a modification of the reference method protocol was tested. After a primary enrichment in UVM for 24h, samples were plated to RAPID' *Listeria* spp, for results 24h sooner than the reference method. There was no significant difference between the 48h reference method and the 24h RAPID' *Listeria* spp method. Routine testing of the food processing environment for the presence of *Listeria* spp. as part of a Hazard Analysis of Critical Control Point (HACCP) program is a way to ensure the sanitation of the processing plant and of the food it produces. Results 24h sooner can be an early alert that conditions exist that can potentially support the growth of pathogenic organisms, like *Listeria monocytogenes*. In addition, since the RAPID' *Listeria* spp method only uses one broth, as opposed to the two broths that the reference method used, there is a cost savings as well.

Strain	Reference	Origin	Typical colonies
<i>Listeria monocytogenes</i> 1/2 a	L7	Munster (raw milk cheese)	+
<i>Listeria monocytogenes</i> 1/2 a	L10	Potted meat	+
<i>Listeria monocytogenes</i> 1/2 a	L12	Smoked salmon	+
<i>Listeria monocytogenes</i> 1/2 a	L128	Soy bean cattle cake	+
<i>Listeria monocytogenes</i> 1/2 b	L13	Pork ears in jelly	+
<i>Listeria monocytogenes</i> 1/2 b	L37	Maroille (raw milk cheese)	+
<i>Listeria monocytogenes</i> 1/2 b	L51	Germain (raw milk cheese)	+
<i>Listeria monocytogenes</i> 1/2 c	L17	Pork breast	+
<i>Listeria monocytogenes</i> 1/2 c	L18	Munster (raw milk cheese)	+
<i>Listeria monocytogenes</i> 1/2 c	L54	Beef Bourguignon	+
<i>Listeria monocytogenes</i> 1/2 c	L117	Montbéliard sausage	+
<i>Listeria monocytogenes</i> 3 a	L191	Fishery environment	+
<i>Listeria monocytogenes</i> 3 a	L192	Fishery environment	+
<i>Listeria monocytogenes</i> 3 b	L55	SLCC 2540 (human)	+
<i>Listeria monocytogenes</i> 3 b	L193	Fishery environment	+
<i>Listeria monocytogenes</i> 3 c	L56	SLCC 2479	+
<i>Listeria monocytogenes</i> 4 a	L57	ATCC 19114 (ruminant brain)	+
<i>Listeria monocytogenes</i> 4 b	L32	Munster (raw milk cheese)	+
<i>Listeria monocytogenes</i> 4 b	L58	Salad	+
<i>Listeria monocytogenes</i> 4 d	L60	ATCC 19117 (Sheep)	+
<i>Listeria monocytogenes</i> 4 d	L194	Fishery environment	+
<i>Listeria monocytogenes</i> 4 e	L62	Reblochon (raw milk cheese)	+
<i>Listeria monocytogenes</i> 4 e	L63	Munster (raw milk cheese)	+
<i>Listeria monocytogenes</i> 7	L67	SLCC 2482 (human feces)	+
<i>Listeria innocua</i> 6 a	L1	ATCC 33090 (cow brain)	+
<i>Listeria innocua</i> 6 a	L77	Toulouse sausage	+
<i>Listeria innocua</i> 6 b	L76	Ground meat	+
<i>Listeria innocua</i> 6 b	L144	Bin	+
<i>Listeria innocua</i>	L88	Pork sausage	+
<i>Listeria innocua</i>	L175	Process water	+
<i>Listeria ivanovii</i> 5	L151	Ground meat	+
<i>Listeria ivanovii</i> 5	L153	Environment	+
<i>Listeria ivanovii</i> 5	L154	Sausages with herbs	+
<i>Listeria ivanovii</i> 5	L182	Environment	+
<i>Listeria ivanovii</i> 5	L184	Birds trap	+
<i>Listeria welshimeri</i> 6 a	L89	Ground meat	+
<i>Listeria welshimeri</i> 6 b	L90	Ground meat	+
<i>Listeria welshimeri</i> 6 b	L86	ATCC 35897 (decaying plant material)	+
<i>Listeria welshimeri</i>	L91	Dried pork sausage	+
<i>Listeria welshimeri</i>	L101	Ham	+
<i>Listeria welshimeri</i>	L155	Raw salmon fillet	+
<i>Listeria welshimeri</i>	L174	Spinach	+
<i>Listeria seeligeri</i> 1/2 b	L82	ATCC 35897 (soil)	+
<i>Listeria seeligeri</i> 1/2 b	L83	Ox tongue	+
<i>Listeria seeligeri</i> 1/2 b	L84	Ground meat	+
<i>Listeria seeligeri</i>	L115	Lake water sampling	+
<i>Listeria seeligeri</i>	L140	Frozen french fries	+
<i>Listeria seeligeri</i>	L189	Frozen french fries	+
<i>Listeria grayi</i>	L81	ATCC 19120 (animal feces)	+
<i>Listeria grayi</i>	L143	Frozen french fries	-
<i>Listeria grayi</i>	L188	Environment	+

ATCC = American Type Culture Collection, USA

L = *Listeria* culture collection, Institut Pasteur de Lille, FranceSLCC = Seeliger's *Listeria* Culture Collection, Würzburg, Germany

Table 2 – Exclusivity Results (1)

Strain	Reference	Origin	Colony color
<i>Bacillus cereus</i>	IPL-BA1	Raw egg	No growth
<i>Bacillus cereus</i>	IPL-BA2	Beet root	No growth
<i>Bacillus cereus</i>	IPL-BA3	Plant	No growth
<i>Bacillus cereus</i>	IPL-BA9	Dehydrated potatoes	No growth
<i>Bacillus cereus</i>	IPL-BA14	Custard	No growth
<i>Bacillus cereus</i>	IPL-BA15	Custard	No growth
<i>Bacillus cereus</i>	IPL-BA19	Environment	No growth
<i>Bacillus cereus</i>	IPL-BA21	Tabbouleh with poultry	No growth
<i>Bacillus cereus</i>	IPL-BA11778	ATCC 11778	No growth
<i>Bacillus coagulans</i>	IPL-BA7	Milk product	No growth
<i>Bacillus mycoides</i>	IPL-BA6	Environment	No growth
<i>Bacillus mycoides</i>	IPL-BA24	Soil	No growth
<i>Bacillus pumilus</i>	IPL-BA22	Tabbouleh with poultry	No growth
<i>Bacillus sphaericus</i>	IPL-BA5	Meat product	No growth
<i>Bacillus sphaericus</i>	IPL-BA23	Environment	No growth
<i>Bacillus stearothermophilus</i>	IPL-BA4	Milk product	No growth
<i>Brochotrix thermosphacta</i>	IPL-15	Ground meat	No growth
<i>Carnobacterium divergens</i>	IPL-46	Minced beef	No growth
<i>Carnobacterium gallinarum</i>	IPL-47	Ice slush of chicken carcasses	No growth
<i>Carnobacterium piscicola</i>	IPL-48	Raw milk	No growth
<i>Citrobacter braakii</i>	IPL-CIT86	Pork sausage	No growth
<i>Citrobacter freundii</i>	IPL-CIT24	Meat product	No growth
<i>Corynebacterium flavescens</i>	IPL-COR1	ATCC 10340 (cheese)	No growth
<i>Corynebacterium variabile</i>	IPL-COR2	ATCC 15753 (food)	No growth
<i>Escherichia coli</i>	IPL-EC20	Tomatoes	No growth
<i>Escherichia coli</i>	IPL-EC21	Celery with mayonnaise	No growth
<i>Enterobacter cloacae</i>	IPL-ENT76	Milk powder	No growth
<i>Enterococcus faecalis</i>	IPL-E1	Egg product	No growth
<i>Enterococcus faecalis</i>	IPL-E6	ATCC 19433	No growth
<i>Enterococcus faecium</i>	IPL-E2	ATCC 3286	No growth
<i>Enterococcus faecium</i>	IPL-E7	CIP 54.33 (Canned fish)	No growth
<i>Enterococcus faecium</i>	IPL-E9	Taramasalata	No growth
<i>Enterococcus durans</i>	IPL-E8	Meat product	No growth
<i>Enterococcus durans</i>	IPL-E10	Meat product	Light blue *
<i>Enterococcus durans</i>	IPL-E331	RDC 486	No growth
<i>Enterococcus durans</i>	IPL-E332	RDC 487	No growth
<i>Enterococcus durans</i>	IPL-E19432	ATCC 19432	No growth
<i>Erysipelothrix rhusiopathiae</i>	IPL-49	Spleen of pig with endocarditis	No growth
<i>Jonesia denitrificans</i>	IPL139	CIP 55134T	Colorless
<i>Klebsiella pneumoniae</i>	IPL-EN63	Celery	No growth
<i>Klebsiella pneumoniae</i>	IPL-EN68	Vegetable salad	No growth
<i>Kurthia gibsonii</i>	IPL-42	Meat product	No growth
<i>Lactobacillus acidophilus</i>	IPL-Lb2885	RDC 488	No growth
<i>Lactobacillus casei</i>	IPL-L33	Milk product	No growth
<i>Lactobacillus casei</i>	IPL-Lb9595	ATCC 9595	No growth
<i>Lactobacillus bulgaricus</i>	IPL-Lb120	RDC120	No growth
<i>Lactobacillus fermentum</i>	IPL-Lb9338	ATCC 9338	No growth
<i>Lactobacillus lactis</i>	IPL-L54	Emmental cheese	No growth
<i>Lactobacillus paracasei</i>	IPL-L35	Milk product	No growth
<i>Lactobacillus plantarum</i>	IPL-L34	Milk product	No growth
<i>Lactobacillus spp</i>	IPL-Lb11506	ATCC 11506	No growth
<i>Lactococcus lactis</i>	IPL-LL	Milk product	No growth
<i>Lactococcus lactis</i>	IPL-Lc7056	CIP 70.56	No growth
<i>Micrococcus spp</i>	IPL-M1	Environment	No growth
<i>Pediococcus acidilactici</i>	IPL-Pd240	RDC 240	No growth
<i>Pediococcus damnosus</i>	IPL-Pd29358	Beer	No growth
<i>Pediococcus damnosus</i>	IPL-Pd102264	Beer	No growth
<i>Pediococcus pentosaceus</i>	IPL-Pd119	Beer	No growth
<i>Propionibacterium freundenreichii</i>	IPL-43	Swiss cheese	No growth
<i>Proteus mirabilis</i>	IPL-EN45	Poultry	No growth
<i>Pseudomonas putida</i>	IPL-PS87	Fish	No growth
<i>Pseudomonas putida</i>	IPL-PS90	Fish	No growth
<i>Rhodococcus equi</i>	IPL-32	Meat product	Colorless
<i>Rhodococcus equi</i>	IPL-R2	Lung abscess of foal	No growth

<i>Rhodotorula rubra</i>	IPL-Le1	Pastry	Colorless
<i>Saccharomyces cerevisiae</i>	IPL-Le5	Coffee extract	No growth
<i>Salmonella brandenburg</i>	IPL-3	Pâté	No growth
<i>Salmonella typhimurium</i>	IPL-S31	Egg product	No growth
<i>Salmonella virchow</i>	IPL-S33	Cockle	No growth
<i>Staphylococcus aureus</i>	IPL-ST16	Meat product	No growth
<i>Staphylococcus aureus</i>	IPL-ST17	Frozen yogurt	No growth
<i>Staphylococcus cohnii</i>	IPL-ST21	Smoked salmon	No growth
<i>Staphylococcus epidermidis</i>	IPL-ST3	Yogurt	No growth
<i>Staphylococcus epidermidis</i>	IPL-ST20	Smoked salmon	No growth
<i>Streptococcus anginosus</i>	IPL-Str611	CIP 102921T	No growth
<i>Streptococcus anginosus</i>	IPL-Str1068	CIP 105031	No growth
<i>Streptococcus bovis</i>	IPL-E3	Meat product	No growth
<i>Streptococcus bovis</i>	IPL-Str44	RDC 44	No growth
<i>Streptococcus bovis</i>	IPL-Str5623	CIP 56.23	No growth
<i>Streptococcus equinus</i>	IPL-Str1074	CIP 102504T	No growth
<i>Streptococcus intermedius</i>	IPL-Str1201	CIP 103248T	No growth
<i>Streptococcus salivarius</i>	IPL-Str1075	CIP 102505	No growth
<i>Streptococcus salivarius</i>	IPL-Str1115	CIP 53.158	No growth

\* Two colonies of *E. durans* (ref # L-E10) grew on the plate; they were pinpoint and not typical color of *Listeria* spp. This organism would not be confused for *Listeria* spp by the user.

ATCC = American Type Culture Collection, USA

CIP = Collection Institute Pasteur, France

IPL = Culture collection, Institut Pasteur de Lille, France

RDC = Culture collection, Bio-Rad Laboratories, France

**Table 4 – Method Comparison Results (1)**

Surface	Level	Inoculation	# samples	RAPID' <i>Listeria</i> spp positive	Reference positive	Method Agreement	X2
Stainless steel	Control	0	5	0	0	100%	-
(internal)	Low	3.1x10 <sup>3</sup>	20	19	19	100%	-
Stainless steel	Control	0	5	0	0	100%	-
(independent)	Low	2.7x10 <sup>2</sup>	20	19	19	100%	-
Plastic	Control	0	5	0	0	100%	-
	Low	2.3x10 <sup>3</sup>	20	8	8	100%	-
Ceramic	Control	0	5	0	0	100%	-
	Low	2.3x10 <sup>3</sup>	20	5	8	85%	1.33
Sealed concrete	Control	0	5	0	0	100%	-
	Low	8.7x10 <sup>2</sup>	20	17	19	90%	0.50

**REFERENCES CITED**

1. Lauer, W., Quiring, C., and Patel, A., Evaluation of the RAPID'*Listeria* spp.: A Medium for Detection of *Listeria* spp. From Selected Environmental Surfaces. AOAC Performance Tested Methods<sup>SM</sup> certification number 080701.
2. *Microbiology Laboratory Guidebook* (October 1, 2004) U.S. Department of Agriculture, Food Safety and Inspection Service, Office of Public Health Science, Chapter 8.05. Online at [www.fsis.usda.gov/PDF/MLG\\_8\\_05.pdf](http://www.fsis.usda.gov/PDF/MLG_8_05.pdf). Accessed May 14, 2007
3. Blackman, I.C., Frank, J.F. (1996) *J Food Prot* **59** (8), 827-831
4. Sacchetti, R., Bianucci, F., Ambrogiani, E. (2003) *New Microbiol* **3**, 269-274